

You may use the information and images contained in this document for non-commercial, personal, or educational purposes only, provided that you (1) do not modify such information and (2) include proper citation. If material is used for other purposes, you must obtain written permission from the author(s) to use the copyrighted material prior to its use.

# Evaluation of Microplastic Loading in Diamondback Terrapin and their Associated Habitats: Sediment Pilot Study

Gabrielle Hammerbach<sup>1,2\*</sup>, Mandi Gordon<sup>2</sup>, Danielle DeChellis<sup>2</sup>, Cynthia Howard<sup>1,2</sup>

<sup>1</sup>University of Houston-Clear Lake, College of Science and Engineering, Houston, Texas

<sup>2</sup>University of Houston-Clear Lake, Environmental Institute of Houston, Houston, Texas

\*Corresponding and presenting author (hammerbach@uhcl.edu)

## Introduction

- Microplastics are a globally emerging contaminant of concern.
- Low-lying coastal marshes serve as sinks and filters for terrestrial waste, putting species using these habitats at risk.
- The Diamondback Terrapin (*Malaclemys terrapin*) is the only endemic estuarine-dwelling turtle in North America and is potentially at risk for microplastic exposure or ingestion.
- An extensive literature search was conducted (Tables 1 and 2) to develop a study design, with the goal of evaluating marsh sediments in estuarine habitats.
- As part of a pilot study conducted in March of 2024, sediment samples were collected and are currently undergoing processing using the following study design.
- Results of this pilot study will refine final sampling and processing protocols implemented in a larger-scale study.

**Table 1** Sources for field sample collection study design.

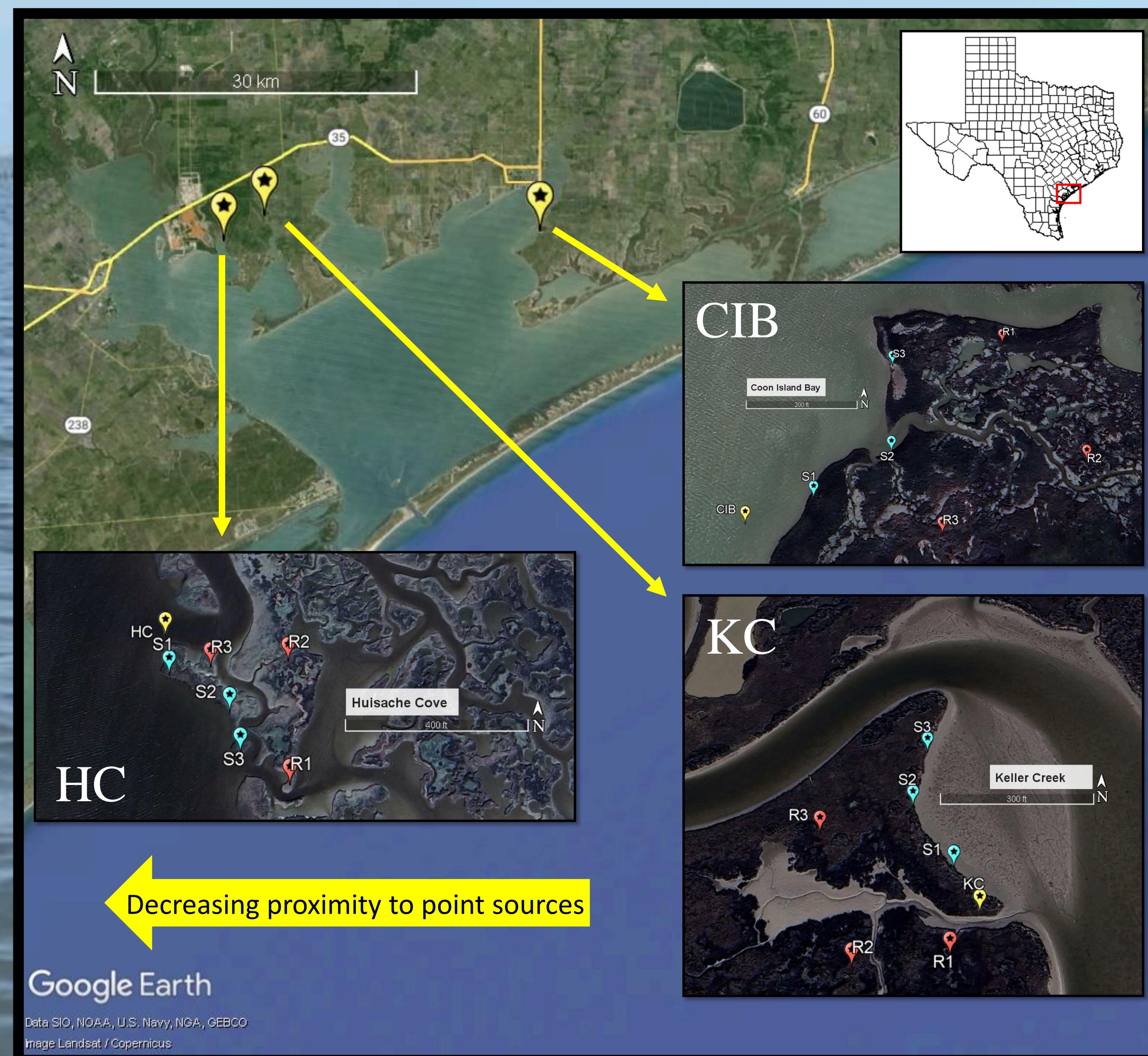
Source	Habitat Type	Core Diameter	Number of Cores	Sample Depth
Alvarez-Zeferino et al. 2020	Beach shoreline	19-cm	10 per site	5-cm
Jenkins and Branco 2020	Salt marsh	9-cm	3 per plot	1-cm
Khan and Prezant 2018	Salt marsh	7.62-cm	3 per plot	10-cm
Klein et al. 2015	Riverine shoreline	Unknown	30-40 per site	2- to 3-cm
Lloret et al. 2021	Estuarine salt marsh	9-cm	3 to 4 per site	2-cm
Pinheiro et al. 2022	Mud flat to high marsh	4.7-cm	6 per zone	5-cm

**Table 2** Sources for sediment processing protocols. Asterisks in Organic Digestion column indicate use of Fenton's reagent.

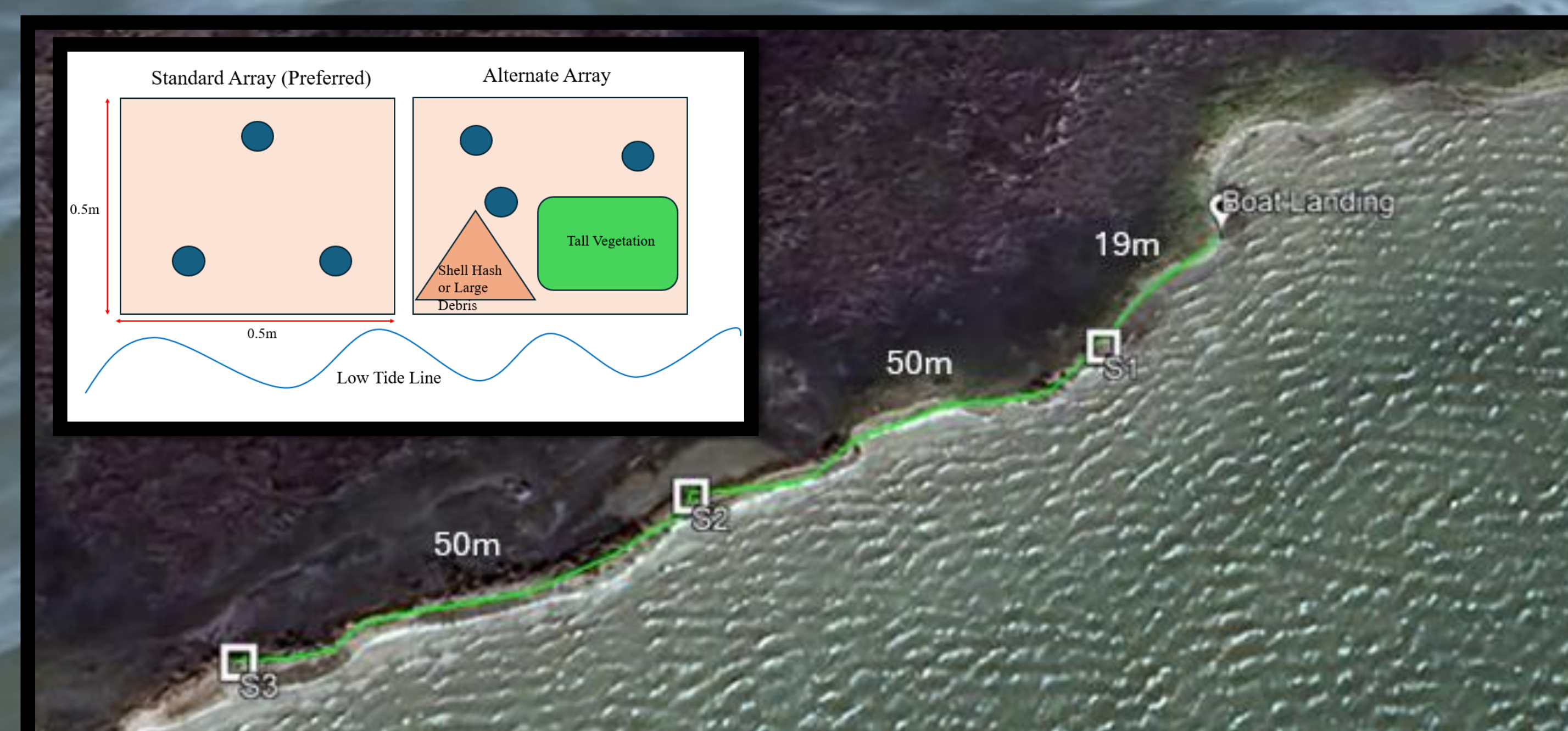
Source	Habitat Type	Sieve Range	Organic Digestion?
Alvarez-Zeferino et al. 2020	Beach shoreline	1.13 to 5-mm	Yes
Beckwith and Fuentes 2018	Beach shoreline	63 to 125- $\mu$ m	No
Hidalgo-Ruz et al. 2012	Beach shoreline	38- $\mu$ m to 4.75-mm	No
Khan and Prezant 2018	Salt marsh	250- $\mu$ m to 4-mm	No
Klein et al. 2015	Riverine shoreline	63 to 630- $\mu$ m	Yes
Lloret et al. 2021	Estuarine salt marsh	250- $\mu$ m to 5-mm	Yes*
Lo et al. 2018	Sandy beaches to mud flats (1:1)	250- $\mu$ m to 5-mm	Yes*
Willis et al. 2017	Riverine estuary	63- $\mu$ m to 4-mm	Yes*
Zhou et al. 2020	Sandy to muddy	5 to 50- $\mu$ m	No

## Pilot Study Field Methods

- Metal and aluminum were used whenever possible to reduce potential for plastic or microplastic contamination; samplers wore brightly colored apparel so fibers could be easily identified and removed during laboratory processing.
- Three sites were selected to assess variation in potential point source microplastic loading, with decreasing proximity to point sources from east to west (Figure 1).
- Two plot types: shoreline and random (Figure 2)
  - The initial shoreline plot located at a randomly determined distance from the vessel and subsequent equidistant-spaced plots were used (3 total shoreline plots per site).
  - The random plot locations were determined by walking 5-10 minutes into the marsh (3 total random plots per site).
- Three 5-cm deep cores were collected from each plot and stored in aluminum tins for transportation back to lab.



**Figure 1** Pilot study field survey area.

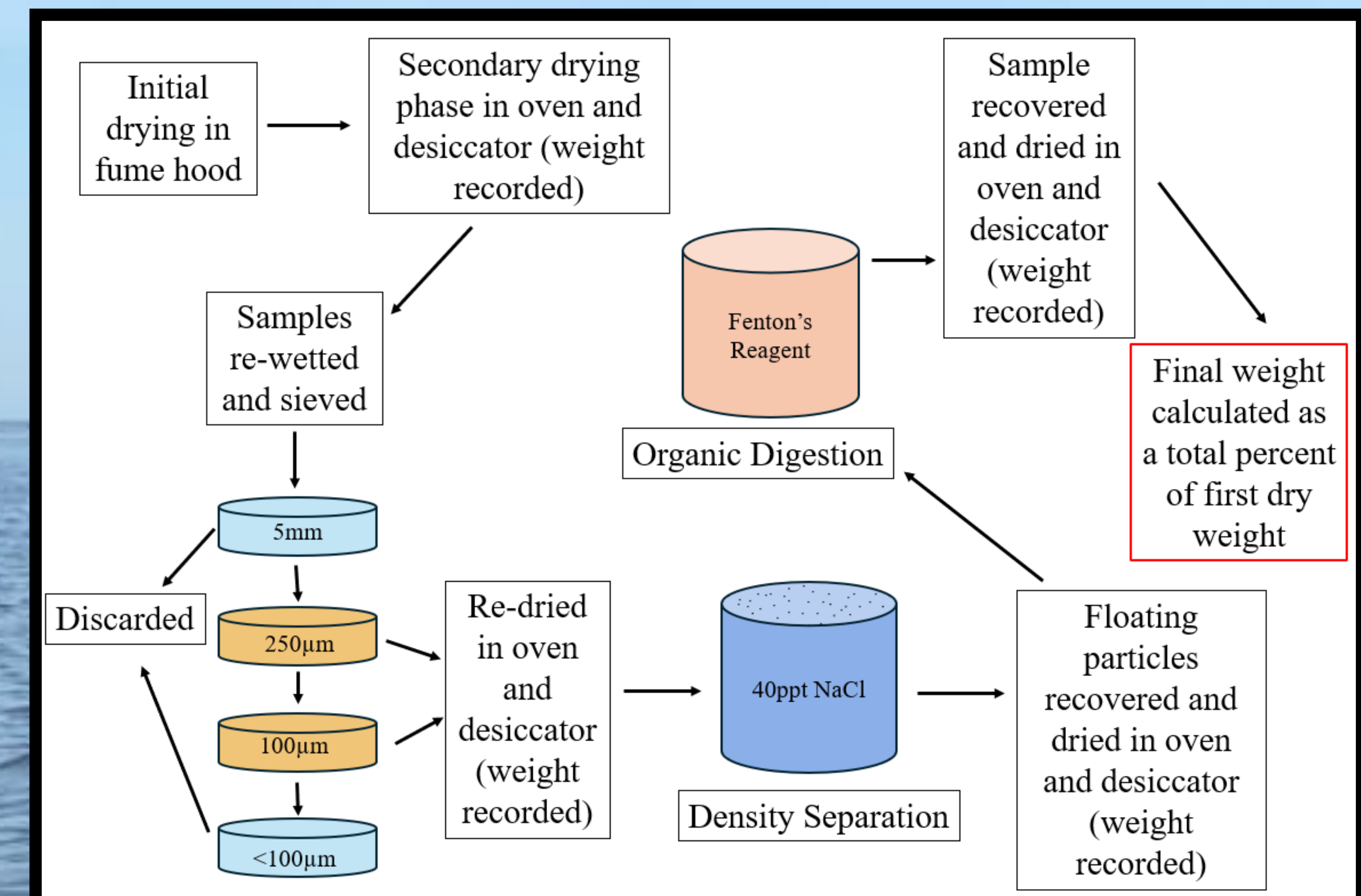


**Figure 2** Shoreline plot and sediment core (inset) distribution.

## Methods

### Laboratory Sample Processing Methods

- Methods adapted from Lloret et al. (2021; Figure 3).
- Samples were initially dried under a fume then placed in an oven at  $< 60^{\circ}\text{C}$ . Drying times and weights were recorded.
- Samples will undergo an initial sieving process through 5-mm, 250- $\mu$ m, and 100- $\mu$ m sized sieves, then dried again.
- Density separation with 40-ppt NaCl solution will be used to remove denser materials followed by another drying phase.
- Organic digestion using Fenton's reagent (Iron (II) and hydrogen peroxide) will be used to remove organic material.
- A final dry weight will be used to calculate the total percentage of the sample that is comprised of microplastics.



**Figure 3** Flowchart of Laboratory Methods

### Future Plans

- Once protocols are refined, additional samples will be collected at new sites in Matagorda and San Antonio Bay as part of a larger study.
- Resulting data will be used to quantify and visualize the gradient of microplastic sediment loading across the bay(s).

### Acknowledgements

- Funding for this work is provided by the Matagorda Bay Mitigation Trust.
- Sampling was conducted under permissions outlined in IACUC protocol #0224.001.R0 and TPWD Scientific Permit for Research #SPR-0504-383



### Literature Cited

Alvarez-Zeferino et al. 2020. Res., Conserv. and Rec. 155:104633.  
 Beckwith et al. 2018. Mar. Poll. Bull. 131(Part A):32-37.  
 Hidalgo-Ruz et al. 2012. Environ. Sci. and Technol. 46:3060-3075.  
 Jenkins et al. 2020. Section IV. In Yozzo et al. pp.1-27.  
 Khan et al. 2018. Mar. Poll. Bull. 130:67-75.  
 Klein et al. 2015. Environ. Sci. and Technol. 49:6070-6076.  
 Lloret et al. 2021. Environ. Advance. 4:100060.  
 Lo et al. 2018. Environ. Poll. 236:208-217.  
 Pinheiro et al. 2022. Sci. of the Tot. Environ. 838:156077.  
 Willis et al. 2017. Frontier. Mar. Sci. 4:419.  
 Zhou et al. 2020. Sci. of the Tot. Environ. 703:134807.

For more information on this project and others at EIH visit our website:

